

# Relative breed contributions to neutral genetic diversity of a comprehensive representation of Iberian native cattle

### J. Cañón<sup>1†</sup>, D. García<sup>1</sup>, J. V. Delgado<sup>2</sup>, S. Dunner<sup>1</sup>, L. Telo da Gama<sup>3</sup>, V. Landi<sup>2</sup>, I. Martín-Burriel<sup>4</sup>, A. Martínez<sup>2</sup>, C. Penedo<sup>5</sup>, C. Rodellar<sup>4</sup>, P. Zaragoza<sup>4</sup> and C. Ginja<sup>3,5</sup>

<sup>1</sup>Departamento de Producción Animal, Facultad de Veterinaria, Universidad Complutense, 28040 Madrid, Spain; <sup>2</sup>Departamento de Genética, Universidad de Córdoba, Campus Rabanales C-5, 14071 Córdoba, Spain; <sup>3</sup>UIRGRMA-INIA, Instituto Nacional dos Recursos Biológicos, Fonte Boa, 2005-048 Vale de Santarém, Portugal; <sup>4</sup>Laboratorio de Genética Bioquímica, Facultad de Veterinaria, Universidad de Zaragoza, Miguel Servet 177, 50013 Zaragoza, Spain; <sup>5</sup>Veterinary Genetics Laboratory, University of California, One Shields Avenue, Davis, CA 95616, USA

(Received 10 September 2010; Accepted 12 January 2011; First published online 11 March 2011)

This study is aimed at establishing priorities for the optimal conservation of genetic diversity among a comprehensive group of 40 cattle breeds from the Iberian Peninsula. Different sets of breed contributions to diversity were obtained with several methods that differ in the relative weight attributed to the within- and between-breed components of the genetic variation. The contributions to the Weitzman diversity and the expected heterozygosity (He) account for between- and within-breed variation only, respectively. Contributions to the core set obtained for several kinship matrices, incorporate both sources of variation, as well as the combined contributions of Ollivier and Foulley and those of Caballero and Toro. In general, breeds that ranked high in the different core set applications also ranked high in the contribution to the global He, for example, Sayaguesa, Retinta, Monchina, Berrenda en Colorado or Marismeña. As expected, the Weitzman method prioritised breeds with low contributions to the He, like Mallorguina, Menorguina, Berrenda en Negro, Mostrenca, Vaca Palmera or Mirandesa, all showing highly negative contributions to He – that is, their removal would significantly increase the average He. Weighing the within- and between-breed components with the F<sub>ST</sub> produced a balanced set of contributions in which all the breeds ranking high in both approaches show up. Unlike the other methods, the contributions to the diversity proposed by Caballero and Toro prioritised a good number of Portuguese breeds (Arouguesa, Barrosã, Mertolenga and Preta ranking highest), but this might be caused by a sample size effect. Only Sayaguesa ranked high in all the methods tested. Considerations with regard to the conservation scheme should be made before adopting any of these approaches: in situ v. cryoconservation, selection and adaptation within the breeds v. crossbreeding or the creation of synthetic breeds. There is no general consensus with regard to balancing within- and between-breed diversity and the decision of which source to favour will depend on the particular scenario. In addition to the genetic information, other factors, such as geographical, historical, economic, cultural, etc., also need to be considered in the formulation of a conservation plan. All these aspects will ultimately influence the distribution of resources by the decision-makers.

Keywords: Iberian cattle, diversity, conservation priorities, microsatellites

#### Implications

The extensive collection of Iberian breeds analysed and the many different methods used to rank them in their contribution to the global genetic diversity makes this study a useful tool for authorities and decision-makers on a local, national or international scale. The results can be combined with other sources of information, such as geographical, economic, historical or cultural, to establish priorities and shares in the allocation of economic resources for diversity

#### Introduction

The erosion of genetic diversity in many local livestock breeds as a consequence of random genetic drift has become one of the main concerns for animal breeders. In this study, most, if not all, of the breeds included have not suffered any relevant directional selection because of different

conservation. In addition, this data set can be applied in a large-scale analysis of the European cattle, which would probably reveal interesting results about the preservation of peripheral and more variable breeds.

<sup>&</sup>lt;sup>+</sup> E-mail: jcanon@vet.ucm.es

circumstances: reduced rate of artificial insemination, reduced effective population sizes, which mean low genetic variability to select from, and selection programmes which, when they exist, are not well organised and have been recently implemented (less than four generations). The reduction of genetic diversity can be observed directly from the loss of heterozygosity (He) and of allelic variants, and neutral molecular markers can be used to assess the levels of genetic variation.

The domestication process of taurine cattle (Bos taurus taurus) has driven the formation of subpopulations (breeds; Scherf, 2000), and genetic drift converted part of the original genetic variation into genetic variation between breeds. As this process of subdivision allows for genetic differentiation between breeds, and although loss of genetic diversity occurs faster in small populations, the structure of a species subdivided into genetically isolated breeds can be, under certain conditions, a good strategy to maintain overall genetic variability (Wright, 1951). However, many European cattle breeds are in danger of becoming extinct (Scherf, 2000), and are losing genetic variability accumulated during the differentiation process. In this situation, breed conservation can contribute to prevent global loss of diversity and the combination of within and between breed neutral genetic diversity can be used to prioritise genetic resources for conservation purposes (Eding and Meuwissen, 2001).

The Iberian Peninsula constitutes a geographical unit and, for long periods of time, its two countries – Portugal and Spain – also formed a political unit. The permeability along the border contributed to the history of the formation of many of the cattle breeds from this region (Sánchez-Belda, 1984). Both countries are characterised by a large number of local cattle breeds, most of which are considered threatened genetic resources. Although many of the breeds included in this study have been previously characterised genetically to some extent together with other breeds (Martín-Burriel *et al.*, 1999; Cañon *et al.*, 2001; Mateus *et al.*, 2004; Cymbron *et al.*, 2005; Martín-Burriel *et al.*, 2007; Ginja *et al.*, 2010a), a comprehensive analysis of the genetic diversity present in the Iberian Peninsula and a source of important bovine genetic resources has never been made.

Under the assumption that the main genetic force acting is genetic drift, with the well-known consequences of increasing both within-population homozygosity and betweenpopulations genetic differences (Falconer and Mackay, 1996), neutral markers, such as microsatellites, are the most appropriate to evaluate the relative contribution of each breed to overall genetic diversity (Boettcher *et al.*, 2010). The substantial genetic differentiation and relatively high genetic diversity found in Iberian cattle should allow for a relevant analysis of genetic contributions. The estimation of these contributions is not trivial and does not have a unique solution (Caballero *et al.*, 2010). There is also a conceptual dichotomy with regard to the importance of between-breed, as opposed to within-breed variation.

The objective of this study was to assess the relative importance of each breed in a comprehensive set of 40 out of 52 lberian cattle breeds for maintenance of genetic diversity, by using a variety of methods to compute conservation priorities according to the estimation of breed contributions to the global diversity. The different methods combine within- and between-breed information in several ways, so as to give decision-makers a wide perspective of the distribution of the genetic variation among all breeds and thus to support conservation policies.

#### **Material and methods**

#### Sampling and genotyping

Information about the 40 native breeds analysed in this study is summarised in Table 1. Figure 1 shows breed geographical distribution.

**Table 1** Breed names, acronyms, sample sizes and country of origin for40 Iberian cattle breeds

Breed	Acronym	Sample size	Country
Alistana	ALS	50	Spain
Asturiana de la Montaña	ASM	50	Spain
Asturiana de los Valles <sup>1</sup>	ASV	50	Spain
Avileña <sup>1</sup>	AVI	50	Spain
Berrenda en Colorado	BC	40	Spain
Berrenda en Negro	BN	30	Spain
Betizu	BET	20	Spain
Bruna dels Pirineus	BRP	50	Spain
Lidia <sup>1</sup>	TL	50	Spain
Mallorquina	MAL	50	Spain
Marismeña	MA	50	Spain
Menorquina	MEN	50	Spain
Monchina	MON	50	Spain
Morucha <sup>1</sup>	MOR	50	Spain
Mostrenca	MOS	50	Spain
Negra Andaluza	NAN	21	Spain
Pajuna	PAJ	38	Spain
Parda de Montaña	PM	50	Spain
Pasiega	PAS	50	Spain
Pirenaica <sup>1</sup>	PIR	50	Spain
Retinta <sup>1</sup>	RET	50	Spain
Rubia Gallega <sup>1</sup>	RGA	50	Spain
Sayaguesa	SAY	50	Spain
Serrana de Teruel	STE	50	Spain
Tudanca	TUD	50	Spain
Vaca Canaria	VCA	50	Spain
Vaca Palmera	PAL	50	Spain
Alentejana <sup>1</sup>	ALT	38	Portugal
Arouquesa	ARO	70	Portugal
Barrosã	BAR	69	Portugal
Brava de Lide	BRA	43	Portugal
Cachena	CAC	51	Portugal
Garvonesa	GAR	39	Portugal
Marinhoa	MRI	46	Portugal
Maronesa	MRO	47	Portugal
Mertolenga <sup>1</sup>	MER	64	Portugal
Minhota	MIN	50	Portugal
Mirandesa	MIR	54	Portugal
Preta	PRE	60	Portugal
Ramo Grande- Açores	RG	44	Portugal

<sup>1</sup>Risk status = not endangered according to their respective National authorities.

#### Neutral genetic diversity of Iberian native cattle

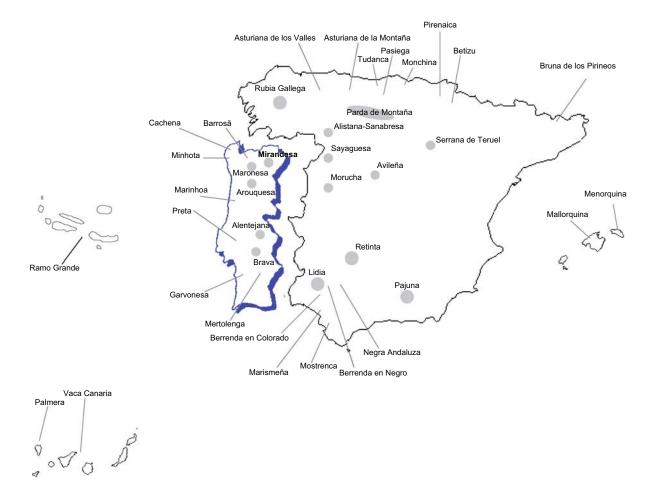


Figure 1 Breed geographical distribution.

About 21 microsatellite loci recommended by the Food and Agriculture Organization (FAO, 2004) for use in genetic diversity studies were analysed: *BM1818, BM1824, BM2113, CSRM60, CSSM66, ETH3, ETH10, ETH185, ETH225, HAUT27, HEL9, ILSTSO06, INRA032, INRA035, INRA037, INRA063, MM12, SPS115, TGLA53, TGLA122* and *TGLA227.* A total of 30 samples, representing the entire allele ranges for this set of markers, were genotyped by all participating laboratories to assure compatibility of results from different equipment and laboratories. Data for *INRA035* and *INRA037* were removed due to the presence of null and 1 bp insertion/deletion alleles, respectively.

Further details on sample collection, DNA extraction and genotyping procedures can be found in previous publications (Martín-Burriel *et al.*, 1999; Cañon *et al.*, 2001; Ginja *et al.*, 2010a).

#### Statistical analysis

First, the Weitzman (1992) approach was applied to calculate the partial contributions (PC) of each breed to the total genetic diversity. The Reynolds genetic distance (Reynolds *et al.*, 1983) was used in the algorithm. Given the large number of breeds involved, the application of Weitzman's formula is too computationally intensive and not practical. Therefore, an approximation of the algorithm must be used. The method proposed by Garcia *et al.* (2005) was applied here. Several thresholds, ranging from 0.4 to 0.25, were used to test the speed of the approximation method and verify the coherence of the results. The threshold corresponds to the coefficient of variation (CV) of the diversity matrix, so that, for each iteration, if the CV is lower than the threshold, the approximation is launched; otherwise the exact algorithm is applied. Therefore, the lower the threshold, the more accurate the results will be, at the expense of computation time. A threshold of 0.25 was finally used to assess breed contributions to the overall diversity. As the approximate and the real results cannot be directly compared, the different thresholds were compared to study their behaviour as the threshold became tighter.

The absence of within-unit information in the Weitzman approach is evident, and has been criticised as an important flaw of this diversity measure (Caballero and Toro, 2002; Eding *et al.*, 2002). Alternatives to include that information in the final contributions have been suggested (Garcia *et al.*, 2005; Ollivier and Foulley, 2005). The aggregate diversity of Ollivier and Foulley (2005) was used here, but the approach of Garcia *et al.* (2005) could not be used because accurate effective population sizes were not available.

The Core Set method (Eding *et al.*, 2002) was also applied to take into account within-breed information. Given the absence of genealogical data to calculate within- and

between-breed kinships, molecular estimations were used: marker estimated kinships (MEK; Eding and Meuwissen, 2001), its extensions based on log-linear regressions, obtained with the weighted log-linear model (WLM) and weighted log-linear mixed model (WLMM; Eding and Meuwissen, 2003) and average molecular coancestries ( $f_m$ ) based on allele frequencies (Caballero and Toro, 2002). The kinship distance was used to analyse breed relationships in a hierarchical Neighbour-joining (NJ) tree and as a basis for their order in contour plots of the kinship coefficients.

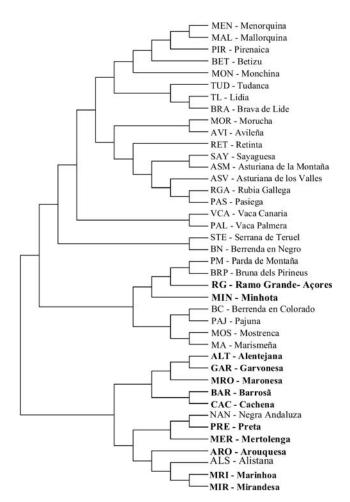
Strict within-breed diversity contributions were assessed through the individual contributions of each breed to the total heterozygosis and calculated as the proportion of global He lost (or gained) from the removal of each breed ( $PC_{He}$ ). Within- and between-breed information was combined according to Ollivier and Foulley (2005).  $F_{ST}$  was calculated to weigh the linear combination of the within- and between-breed PC<sub>FST</sub>. Alternatively, following Chaiwong and Kinghorn (1999) and Piyasatian and Kinghorn (2003), more weight was given to the between-breed component, up to a 5:1 ratio ( $PC_{5:1}$ ).

Caballero and Toro (2002) and Fabuel et al. (2004), proposed another method for combining within- and betweenbreed information. Using the same rationale behind the Core Set method, which defines diversity as the opposite of average coancestry, they derived a partition of the coancestry into within- and between-breed components, which can be calculated for each breed using the average within-breed coancestry and the average minimum Nei's distance between that breed and all the others. One important drawback of this approach is that each breed's contribution is weighted by the fraction of the global population size corresponding to that breed, that is, population sizes are needed for all the breeds. In the absence of this information, sample sizes can be used instead, and the more the ratio of the sample size of each breed relative to the total sample size resembles the equivalent population ratio, the more accurate the estimates will be. These estimates were also calculated ignoring sample sizes, as suggested in Fabuel et al. (2004).

#### Results

#### Breed relationships

The NJ tree obtained for the 40 breeds using the kinship distance is shown in Figure 2. The clustering pattern fits the regional location of the breeds with some exceptions. The two major clusters correspond roughly to the country of origin. Only three Portuguese breeds clustered out of their group. Brava de Lide and De Lidia were separated from the rest due to the fact that both breeds are related and both share a history of genetic isolation from other breeds (Cañon *et al.*, 2008; Ginja *et al.*, 2010a). Crossbreeding with other breeds not represented in the data set might explain Ramo Grande (Azores) and Minhota clustering together with Bruna de los Pirineos and Parda de Montaña, although it makes sense that the latter two breeds are close together, as they are closely located geographically. Pirenaica inexplicably



**Figure 2** Neighbour-joining tree computed with the kinship distance. Breeds from Portugal are shown in bold.

grouped with the two breeds from the Balearic Islands, Mallorquina and Menorquina.

Breeds from Northern Spain (Rubia Gallega, Asturiana de los Valles, Asturiana de la Montaña, Pasiega and Sayaguesa) clustered together accordingly and in close genetic relationship with a cluster of breeds from Western–Central Spain (Morucha, Avileña and Retinta).

One would expect Negra Andaluza to group with the Central–Southern Spanish breeds, but instead it is embedded in the Portuguese cluster, close to the Preta breed, which is distributed in Central Portugal around the Tagus River. Although this result could be due to a bias from the lower sample size of the Negra Andaluza, it might also reflect a common history between these two breeds. The other Spanish breed in the Portuguese cluster is Alistana, which has a shared origin with Mirandesa.

## *Breed contributions to overall diversity – Weitzman algorithm*

Figure 3 shows the evolution of breed contributions in percentage along the different thresholds.

The results show a group of breeds (Mallorquina, Palmera, Mirandesa, Menorquina, Berrenda en Negro and Mostrenca)

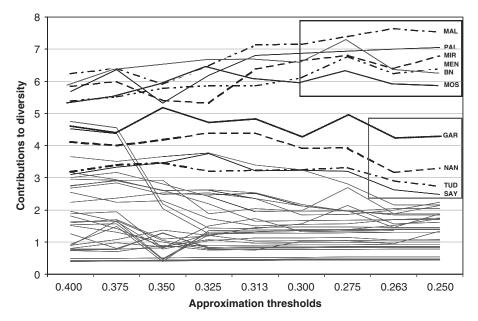


Figure 3 Evolution of breed contributions (%) along the different thresholds tested (MAL = Mallorquina; PAL = Palmera; MIR = Mirandesa; MEN = Menorquina; BN = Berrenda en Negro; MOS = Mostrenca; GAR = Garvonesa; NAN = Negra Andaluza; TUD = Tudanca; SAY = Sayaguesa).

Table 2 Pairwise	correlation	coefficients	between l	breed	contributions t	o We	itzman's diversi	ity
------------------	-------------	--------------	-----------	-------	-----------------	------	------------------	-----

Threshold	0.4	0.375	0.35	0.325	0.31275	0.3	0.275	0.2625
0.4								
0.375	0.9902							
0.35	0.9257	0.9341						
0.325	0.8976	0.9177	0.9715					
0.31275	0.8965	0.9177	0.9567	0.9901				
0.3	0.8962	0.9185	0.9499	0.9822	0.9955			
0.275	0.9022	0.9238	0.9522	0.9771	0.9884	0.9948		
0.2625	0.8819	0.9065	0.9260	0.9637	0.9830	0.9909	0.9910	
0.25	0.8851	0.9082	0.9257	0.9593	0.9805	0.9893	0.9904	0.9978

that consistently had a higher level of contribution and could be prioritised for conservation according to Weitzman's diversity (black box). A second breed group (Garvonesa, Negra Andaluza, Tudanca and Sayaguesa), with contributions ranging from 2.5% to almost 4% at the 0.25 threshold, could be considered as having a second level of conservation priority (grey box). The remaining 30 breeds had lower contributions (<2.5%, unboxed) to diversity. To assess the consistency of these results, pairwise correlation coefficients were calculated between breed contributions for each pair of thresholds. As shown in Table 2, these were high enough (>0.88 between the two extreme thresholds, >0.99 between consecutive thresholds from 0.35 to 0.25) to consider the results consistent.

## Partitioning of the genetic diversity into within- and between-breed components

As described in the 'Material and methods' section, different procedures were applied to estimate within- and betweenbreed kinships in order to obtain breed contributions to the core set. Figure 4 shows contour plots of the MEKs obtained from individual genotypes (top) and from average coancestries calculated with allelic frequencies (bottom). In these plots, redder areas indicate higher kinship values. The breeds were arranged in the same order as in the NJ tree of Figure 2 for a better view of the clusters. The two clusters formed by Brava de Lide and Lidia and by Mallorguina and Menorguina were the most inbred. Vaca Palmera, Berrenda en Negro and Garvonesa also showed higher levels of within-breed kinship. With the possible exception of the Brava/Lidia pair, the Mostrenca breed not only had high within-breed kinship, but was more related than any other breed, to the Portuguese breed group of Alentejana, Garvonesa, Maronesa, Barrosã and Cachena, which clustered together very clearly. A certain level of relatedness was also detected among this Portuquese breed group and the Arouquesa, Mertolenga, Preta and Negra Andaluza breeds. Two other breed clusters were confirmed by the Kinship analysis: the Alistana/ Mirandesa/ Marinhoa and the Berrenda en Negro/Parda de Montana/ Bruna de los Pirineos/Ramo Grande/Minhota groups. Contrary to expectations that breeds with high levels of withinbreed kinships such as Brava, Lidia or Mallorquina, would

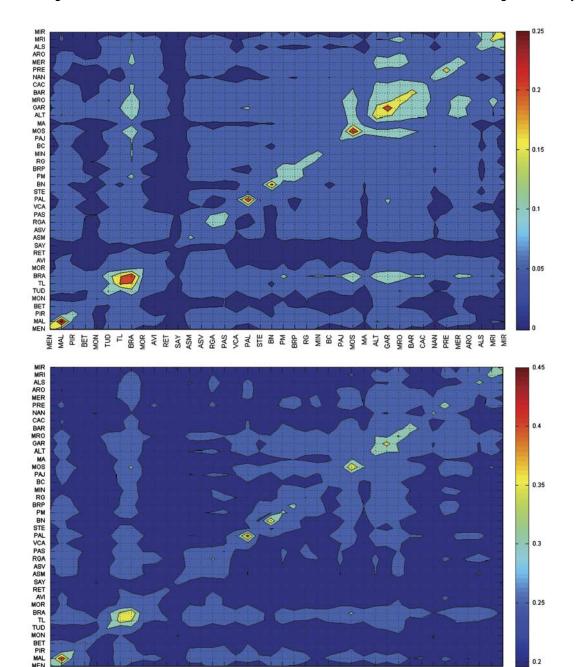


Figure 4 Contour plots of MEKs (marker estimated kinships; Eding and Meuwissen, 2001; top) and average coancestries, f<sub>m</sub> (bottom).

show low between-population values, their between-breed kinships were remarkably uniform and moderately high with all other breeds, especially when allelic frequencies were used in the calculations.

NON UT

AOR

AVI SAY SAY ASM ASM ASV ASV RGA PAS PAS

MAL

Table 3 shows breed contributions to the core set according to the different kinship matrices (Core MEK, Core  $f_{\rm m}$ , Core WLM and Core WLMM), Weitzman's diversity with a 0.25 threshold for the approximation (PC<sub>We0.25</sub>), expected Hes (PC<sub>He</sub>) and linear combinations of the latter two approaches, using F<sub>ST</sub> and (1-F<sub>ST</sub>) to weight the betweenand within-breed contributions, respectively (F<sub>ST</sub> = 0.0898), and alternatively a 5:1 ratio favouring the between breed component ( $PC_{FST}$  and  $PC_{5:1}$ , respectively). A disadvantage of the Core Set method is that sampling errors or populational effects can lead to negative contributions to the core set (Eding *et al.*, 2002). As a solution, the authors proposed to iteratively make the contribution of the breed with the lowest value (most negative) zero and recalculate all contributions excluding that breed. Eventually, the algorithm will reach a suboptimal solution where there are no negative contributions. In this case, the solution was reached after zeroing 32 breeds, so that only eight breeds showed positive values. Similarly, only 12 and 10 breeds showed positive values with the average coancestry and WLM matrices,

**Table 3** Breed contributions to the core set using different kinship matrices, to the Weitzman diversity, to the global expected He and linear combinations of the latter two approaches, using  $F_{ST}$  and a 5:1 ratio to weight between- and within-breed diversity. Highly prioritised breeds are bold and shaded dark and moderately prioritised breeds are shaded light

Breed	Core MEK	Core <i>f</i> m	Core WLM	Core WLMM	PC <sub>We0.25</sub>	PC <sub>He</sub>	PC <sub>Fst</sub> (0.0898)	PC <sub>5:1</sub> (0.83)
BET	0.051	0.014	0.004	0.032	2.232	0.173	0.357	1.889
TL	0	0	0	0.022	1.621	-0.111	0.045	1.332
MEN	0.007	0.02	0.026	0.028	6.401	-0.102	0.482	5.317
ALS	0	0	0	0.028	1.701	0.067	0.214	1.429
SAY	0.318	0.263	0.212	0.061	2.474	0.251	0.451	2.103
TUD	0	0	0	0.026	2.722	0.093	0.329	2.284
ASV	0	0	0	0.03	0.541	0.244	0.27	0.491
ASM	0	0	0	0.023	1.063	0.187	0.266	0.917
RET	0.153	0.097	0.178	0.047	1.456	0.265	0.372	1.258
MOR	0	0	0	0.029	0.838	0.211	0.267	0.734
AVI	0	0	0	0.032	1.07	0.186	0.266	0.923
PIR	0	0	0	0.025	1.848	0.139	0.292	1.563
RGA	0	0	0	0.021	0.992	0.112	0.191	0.846
MAL	0	0	0	0.013	7.523	-0.392	0.319	6.204
MON	0.131	0.22	0.037	0.04	1.879	0.219	0.368	1.602
STE	0	0	0	0.031	0.925	0.269	0.328	0.816
PM	0	0	0	0.015	0.46	0.132	0.161	0.405
BRP	0	0	0	0.015	1.332	0.132	0.24	1.132
PAS	0	0	0	0.021	0.871	0.137	0.203	0.749
BC	0	0.045	0.328	0.054	0.475	0.197	0.222	0.429
BN	0	0	0	0.028	6.325	-0.197	0.389	5.238
MOS	0	0	0	0	5.916	-0.276	0.28	4.884
PAJ	0	0	0	0.024	0.783	0.161	0.217	0.679
NAN	0	0.129	0.048	0.043	3.291	0.08	0.368	2.756
MA	0.164	0.017	0.032	0.042	1.383	0.192	0.299	1.184
VCA	0.089	0.07	0.099	0.044	1.728	0.205	0.342	1.474
PAL	0	0	0	0.02	7.048	-0.335	0.328	5.818
ALT	0	0	0	0.006	1.741	-0.219	-0.043	1.414
ARO	0	0	0	0.02	0.866	0.02	0.096	0.725
BAR	0	0	0	0.018	1.88	-0.182	0.003	1.536
BRA	0	0	0	0.001	1.305	-0.431	-0.275	1.015
CAC	0	0.015	0	0.021	0.442	-0.027	0.015	0.364
GAR	0	0	0	0.009	4.236	-0.356	0.056	3.47
MRI	0	0	0	0.02	2.131	-0.122	0.08	1.755
MRO	0	0	0	0.007	2.043	-0.281	-0.073	1.656
MER	0	0	0	0.016	1.302	-0.057	0.065	1.075
MIN	0	0.028	0	0.021	1.409	-0.015	0.113	1.172
MIR	0.088	0.081	0.037	0.032	6.798	-0.344	0.298	5.608
PRE	0	0	0	0.016	2.154	-0.187	0.023	1.764
RG	0	0	0	0.017	1.356	-0.035	0.089	1.124

 $He = heterozygosity; MEK = marker estimated kinships; f_m = average molecular coancestries; WLM = weighted log-linear model; WLMM = weighted log-linear mixed model; PC = partial contributions.$ 

respectively. Highly prioritised breeds are easily identified, but there is no way to discriminate among breeds with null contributions. The WLMM was developed to prevent this situation, and adjusts the matrix so that there can be as few as only one breed contributing to zero.

Sayaguesa consistently appeared as the breed contributing most in three of the four core set applications, and in the fourth it ranked second. Furthermore, its contribution to the Weitzman diversity, ranked in the top 10. Retinta was among the top three and Monchina ranked second, fourth and seventh in three of the four Core Set methods. Berrenda en Colorado had the highest contribution to the core set according to the WLM method, and ranked second with the WLMM, but had little contribution using the average coancestries and none with the plain MEKs. Similarly, Marismeña had the second largest contribution using MEKs and ranked sixth with the WLMM model, but had little contributions under the WLM model and with the average coancestry matrix. Negra Andaluza and Vaca Canaria also had noticeable contributions to the different core sets, with Negra Andaluza ranking second

Breed	f <sub>ii</sub>	DNei	Contribution to $\bar{f}$	Contribution to D	Proportional contribution (%)	Proportional contribution ignoring sample size (%)	
BET	0.311	0.077	0.0024	0.008	1.056	2.541	
TL	0.383	0.091	0.0076	0.018	2.438	2.347	
MEN	0.371	0.107	0.0068	0.019	2.537	2.442	
ALS	0.316	0.075	0.0063	0.020	2.612	2.514	
SAY	0.274	0.078	0.0051	0.021	2.769	2.665	
TUD	0.312	0.079	0.0061	0.020	2.640	2.541	
ASV	0.271	0.052	0.0057	0.020	2.688	2.588	
ASM	0.292	0.060	0.0060	0.020	2.646	2.547	
RET	0.273	0.064	0.0054	0.021	2.727	2.625	
MOR	0.278	0.059	0.0057	0.020	2.687	2.586	
AVI	0.286	0.065	0.0057	0.020	2.685	2.584	
PIR	0.308	0.072	0.0061	0.020	2.629	2.531	
RGA	0.306	0.065	0.0063	0.020	2.611	2.514	
MAL	0.454	0.125	0.0085	0.017	2.311	2.224	
MON	0.275	0.074	0.0052	0.021	2.750	2.647	
STE	0.271	0.055	0.0056	0.020	2.701	2.599	
PM	0.306	0.062	0.0063	0.020	2.603	2.505	
BRP	0.310	0.067	0.0063	0.020	2.608	2.510	
PAS	0.302	0.065	0.0062	0.020	2.627	2.529	
BC	0.259	0.051	0.0043	0.016	2.181	2.624	
BN	0.401	0.118	0.0044	0.011	1.480	2.375	
MOS	0.401	0.101	0.0078	0.018	2.412	2.322	
PAJ	0.295	0.057	0.0047	0.015	1.996	2.527	
NAN	0.314	0.087	0.0025	0.008	1.118	2.562	
MA	0.289	0.063	0.0059	0.020	2.664	2.564	
VCA	0.283	0.064	0.0057	0.020	2.689	2.589	
PAL	0.424	0.120	0.0079	0.018	2.396	2.307	
ALT	0.340	0.071	0.0053	0.014	1.914	2.424	
ARO	0.277	0.051	0.0082	0.028	3.733	2.567	
BAR	0.326	0.077	0.0089	0.027	3.569	2.489	
BRA	0.397	0.089	0.0069	0.015	2.047	2.292	
CAC	0.289	0.061	0.0060	0.020	2.712	2.559	
GAR	0.375	0.091	0.0057	0.015	1.925	2.376	
MRI	0.325	0.078	0.0059	0.018	2.386	2.496	
MRO	0.357	0.073	0.0070	0.017	2.315	2.371	
MER	0.297	0.06	0.0079	0.025	3.361	2.528	
MIN	0.289	0.062	0.0059	0.020	2.664	2.564	
MIR	0.389	0.123	0.0075	0.021	2.727	2.430	
PRE	0.336	0.080	0.0080	0.023	3.073	2.465	
RG	0.303	0.067	0.0054	0.017	2.316	2.533	

**Table 4** For each breed,  $f_{ii}$ , DNei with rest of the breeds, absolute contribution to the global coancestry, absolute contribution to diversity, proportional contribution to gene diversity and proportional contribution to diversity ignoring sample sizes are shown. Highly prioritised breeds are bold and shaded dark and moderately prioritised breeds are shaded light

 $f_{ii}$  = average within-breed coancestry; DNei = average Nei-distance;  $\bar{f}$  = global coancestry.

when using average coancestries and fifth with the WLMM, where Vaca Canaria was fourth. This breed also had positive contributions in all the other cases, but not much higher than half the contribution of the next higher contributing breed. The WLMM model allowed for comparison among all the 40 breeds, so that a true ranking could be established. In this case, a second group of moderately contributing breeds could be distinguished that included Betizu, Asturiana de Valles, Avileña, Serrana de Teruel and Mirandesa.

As expected due to the nature of the method, Weitzman contributions prioritised breeds that were not particularly favoured by the Core Set methods. Of the top five breeds (Mallorquina, Vaca Palmera, Mirandesa, Menorquina and Berrenda en Negro), only Menorquina and Mirandesa had positive contributions to the first three core sets, and these were rather low.

Contributions to the overall expected He set Sayaguesa again as one of the most prioritised, ranking third, with Serrana de Teruel and Retinta above it. Mochina and Asturiana de los Valles also appeared highly ranked. One of the main drawbacks of calculating the loss of He when removing a breed from the pool is that there will always be many breeds

	Core MEKs	Core ParMed	Core WLM	Core WLMM	PC <sub>We0.25</sub>	PC <sub>He</sub>	PC <sub>fst</sub>	PC <sub>5:1</sub>	Proportional contribution
Core <i>f</i> <sub>m</sub>	0.810								
Core WLM	0.546	0.575							
Core WLMM	0.666	0.682	0.720						
PC	0.015	0.052	-0.096	-0.132					
PC <sub>He</sub>	0.331	0.306	0.339	0.674	-0.635				
PC <sub>FST</sub>	0.417	0.428	0.301	0.665	0.362	0.490			
PC <sub>5:1</sub>	0.022	0.060	-0.090	-0.119	1.000	-0.622	0.378		
Proportional contribution	0.109	-0.019	-0.027	-0.015	-0.264	0.131	-0.140	-0.265	
Proportional contribution ignoring sample size	0.399	0.440	0.413	0.700	-0.652	0.892	0.340	-0.642	0.263

 Table 5
 Pairwise correlation coefficients between the contributions obtained with the different methods, in the same order as shown in Table 3 and Table 4

MEK = marker estimated kinships; WLM = weighted log-linear model; WLMM = weighted log-linear mixed model; PC = partial contributions;  $f_m = average molecular coancestries.$ 

with negative contributions, as all contributions must sum zero. Thus, the removal of a breed actually results in a *gain* in average He. According to this approach, all Portuguese breeds showed negative values, except for Arouquesa that had a very low but positive contribution.

The linear combination of the Weitzman (between-breed component) and the He (within-breed component) contributions, when weighted by the F<sub>ST</sub> generated more balanced results than each of these estimates alone. It ranked the breeds prioritised under both the Core Set and the Weitzman methods as high, with the exception of Berrenda en Colorado and Marismeña. Breeds favoured in the average He approach also ranked high with the linear combination, except Asturiana de los Valles. This combination of diversity components therefore represents a balanced intermediate ranking of breed contributions to overall diversity that incorporates most of the important breeds under either within- or between-breed approaches alone. The 5:1 ratio, however, does not really provide any additional information to that observed with the Weitzman contributions alone. Indeed, the correlation between them is virtually one (>0.999).

The results of the calculations of breed contributions following Caballero and Toro (2002) and the estimates obtained when sample size was ignored (Fabuel et al., 2004) are shown in Table 4. Contrary to the results shown in Table 3, the most prioritised breeds were the Portuguese, with Arouquesa, Barrosã, Mertolenga and Preta as the top four. Sayaguesa ranked as the fifth most contributive breed. Retinta, Monchina and Serrana de Teruel ranked moderately high here too, as they did before. However, these results are only valid if the sample structure reflects the real population structure, that is, if the ratio of breed sample size to total sample size approaches the ratio of breed population size to total population size. In this case, and because most breed sample sizes were similar, those few breeds with larger samples could be favoured, as appears to be the case of the four top ranking breeds.

The contributions obtained ignoring the sample sizes were more uniform, with only slight differences among breeds, which is not very useful for practical prioritisation purposes. It is also arguable whether ignoring sample sizes is much better than using them to obtain biased contributions (see Fabuel *et al.*, 2004 for additional discussion on corrections in the weight).

The linear correlation coefficients calculated for pairwise contributions obtained with the different prioritisation methods are shown in Table 5. As expected, there is a strong negative correlation between the Weitzman and the He contributions (-0.635), and the correlations of the PC<sub>5:1</sub> with all other methods are almost equal to those of the Weitzman approach, because of the excess weighting given to the between-breed component (i.e. Weitzman) in the linear PC<sub>5:1</sub> combination. Correlations with the Core Set methods are not too meaningful, because of the many null contributions. Only the values obtained with the WLMM matrix can lead to some conclusions, like the relatively high correlation of these with the contributions to the global He.

The proportional contributions from Table 4 do not correlate much with those from other methods, except for a moderate negative correlation with the Weitzman and the  $PC_{5:1}$  contributions. As for the proportional contribution obtained ignoring sample size, their correlation with other methods was similar to those from the average He contributions.

#### Discussion

Many of the diversity studies published over the past two decades had the objective of estimating genetic differences between breeds and, as a consequence, priorities for conservation were mainly based on genetic distances (Hodges, 1992), thus ignoring genetic variability and its distribution within breeds.

Management of the genetic diversity in subdivided populations and the prioritisation of breeds for optimal allocation of resources, in order to maximise the global variation, have been the subject of several more recent publications (see review by Toro and Caballero, 2005). Both methodological

and field studies have been widely published in recent years (Bennewitz and Meuwissen, 2004; Fabuel *et al.*, 2004; Mateus *et al.*, 2004; Garcia *et al.*, 2005; Bennewitz *et al.*, 2006; European Cattle Genetic Diversity Consortium, 2006; Hayes *et al.*, 2006; Tapio *et al.*, 2006; Boettcher *et al.*, 2010), and yet no consensus has been reached on how to appropriately account for the between- and within-population components of genetic variation (Meuwissen, 2009; Caballero *et al.*, 2010).

The choice between prioritising within- or between-breed diversity is not an easy task. Both components should be taken into account, but how to weight them is not consensual. Favouring within-breed diversity allows for a better response to selection and adaptation, whereas weighting between-breed variation is useful to assess crossbreeding/ admixture and to create synthetic populations (Barker, 1999; Tapio *et al.*, 2006). Inbred populations with low levels of genetic variability are less viable than those with large effective sizes and high He, but combining distant and eventually inbred populations, with different alleles segregating at high frequencies, leads to new populations with a high genetic variability due to the new heterozygotes.

There is a paradox underlying the use of within- v. betweenpopulation components of genetic variation in conservation calculations. If one favours the between-population diversity, as in the Weitzman or the Piyasatian and Kinghorn (2003) methods, distant and more differentiated breeds, such as Menorquina, Mallorquina, Palmera, Berrenda en Negro, Mostrenca, Garvonesa, Negra Andaluza, Mirandesa or Tudanca, are prioritised. Nonetheless, distant breeds have generally undergone strong genetic drift and have high levels of inbreeding, as reflected here by their high coancestries ( $f_{ii} > 0.312$ , Table 4). They can be just a mere source of alleles rarely found in other breeds and not contribute significantly to overall diversity with within-breed variation. Therefore, it could be argued that breeds with large effective sizes should be prioritised instead of these genetically 'exhausted' breeds. However, all of these breeds, except for Mostrenca and Garvonesa, still ranked high when the weight of the between-breed component was reduced by using the  $F_{ST}$ , as proposed by Ollivier and Foulley (2005). Only the Sayaguesa breed was prioritised by all the methods (Tables 3 and 4), and two more breeds (Retinta and Monchina) were prioritised by most but not all of these approaches.

Furthermore, investing money in breeds prioritised under the between-breed diversity criterion would raise yet another tricky situation: preserving breeds with higher betweenpopulation diversities and implementing breeding programmes aimed at increasing their effective sizes while minimising coancestries, would actually reduce global diversity – still under the hypothesis of considering betweenbreed variation only – as their average distance from the remaining breeds would eventually be reduced. The outcome would be 'healthier' breeds (considering, generally speaking, that a breed is healthy when it has features like high effective size, low homozygosity or low-kinship levels, and thus minimises the loss of genetic diversity by time unit), which is good, but less global variation, which is bad. And yet, one cannot help considering that directing financial resources towards the most endangered breeds seems like a sensible thing to do. But in a scenario in which only between-breed diversity was considered, the way to maintain global diversity would be to keep breeds as far apart as possible. One way to accomplish this is to maintain or increase selection and drift effects, leading to more homozygous populations. This would certainly maximise diversity in hypothetical intercrossings, but in practice no funds would be given by any decision-maker to increase inbreeding and it is also unlikely that local breeders would comply with a policy ultimately aimed at creating breed intercrossings.

If the within-breed component is prioritised instead, those breeds with higher heterozygosities (such as Retinta or Asturiana de los Valles in this study), allelic diversities or effective population sizes would be favoured in the allocation of financial resources. But what is the point in investing more resources in breeds or populations that are already not at risk because they have a 'healthy' genetic structure? Furthermore, higher genetic diversity of some breeds can also be a result of admixture and there would be the risk of preserving crossbred populations, such as Monchina, Serrana de Teruel, Vaca Canaria, Minhota and Ramo Grande-Azores (see also Ginja *et al.*, 2010b).

Approaches focusing on minimising coancestry generally result in opposite priorities to those of distance methods, thus favouring less endangered breeds, which are those that actually need less support. This might be useful in cryoconservation programmes aimed at creating a bank of genetic resources in case a natural or human-triggered disaster brought the species to the verge of extinction and artificial reintroduction of genetic material became necessary. For practical in situ conservation plans, though, it is quite unlikely that local breeders would naturally accept the creation of a mixed new population, and thus the approach would have no purpose. In the context of in situ breed conservation, other methods might prove useful, such as that proposed by Fernández et al. (2008) and implemented in METAPOP software (Perez-Figueroa et al., 2009) in which a certain amount of optimal introduction of migrant genes among the different breeds would be allowed to maximise global genetic diversity.

Still, the concept of conservation priorities seems somewhat pointless for purposes of allocation of resources when local domestic breeds are involved and species viability is not threatened. This is all the more so if one considers the otherthan-genetic factors, such as cultural, historical, economical, etc., that are influential, if not capital, for the decision-makers, along with the fact that *in situ* farm conservation is generally preferred. However, it would be interesting to further narrow population subdivision and determine conservation priorities within breeds, instead of (or maybe complementary to) setting priorities at the breed level. Comparing herds, lineages, regions or other pertinent subdivisions could be of practical management interest, at least in the case of traditional or local breeds where significant population substructure exists, for example, Lidia cattle. The lack of field implementations of the methodologies described here (i.e. actual policies with decisions based at least partly on priorities derived from a genetic diversity analysis) could probably be easily overcome if the underlying hypothesis of admixing subunits such as herds, rather than breeds, were to be used as a background concept to maximise diversity.

The special case of the two fighting bull breeds (Toro de Lidia and Brava de Lide) included in this study must be discussed here. These two breeds have similar production systems and breeding traditions, which resulted in their subdivision into genetically differentiated and mostly isolated lineages named 'encastes' (Cañon et al., 2008; Ginja *et al.*, 2010a). Both breeds maintain a level of genetic diversity that is higher than many other European breeds (Cortes et al., 2008; Ginja et al., 2010c). However, neither of these breeds is prioritised by the methods used in this study, with zero contribution to the core set – except for the WLMM model – and negative contributions to the average He. Furthermore, because of the high level of inbreeding found, Brava de Lide had the most negative contribution to the PC<sub>He</sub> ranking among all breeds. The sampling of 50 unrelated individuals per breed selected across independent herds was originally and theoretically designed to estimate genetic distances between breeds (FAO, 1993). However, it is possible that for highly subdivided breeds in which an important proportion of the genetic variability is distributed among herds, this sampling procedure underestimates breed contributions to the total genetic diversity because it does not represent the real population accurately.

Finally, another consideration to be made relates to the type of markers used, as this is an important factor in accounting for the limitations of this kind of conservation study and the reduced impact on real-life applications to breed conservation. First, the markers in this study are assumed to be neutral and mainly influenced by genetic drift. Therefore, other evolutionary processes of interest, such as adaptation and/or selection for economic traits were not accounted for in the calculations. In addition, the number of microsatellite markers used in this and similar studies is somewhat limited. Both problems will most probably be overcome soon with the release of dense marker maps and the establishment of research projects at the level of genomics (Gibbs et al., 2009). Among other advantages, the use of a large number of markers will allow for more precise estimations of coancestries, which are key to several measures of diversity, and for doing conservation analyses at the level of breeds, herds and individuals. As well, it will be possible to consider the effects of both neutral and adaptive genetic variation from distinct genomic regions (Windig and Engelsma, 2010). This volume of genomic information will also allow for the increase in effective population size and the minimisation of withinindividual variation (Wang and Hill, 2000).

In conclusion, this study analysed the between- and withinbreed components of extant genetic diversity of Iberian cattle, and performed a comprehensive comparison among the methods that can be used to rank breeds for conservation. Neutral genetic diversity of Iberian native cattle

This information can be complemented with that of other markers and used by local authorities in conservation programmes. In addition, the data can be applied in a large-scale analysis of European cattle, which could reveal interesting results with regard to the preservation of peripheral and more variable breeds, such as those of native cattle.

#### Acknowledgements

We wish to acknowledge the financial support of the INIA and European Regional Development Fund no. RZ01-002-C2-1, RZ01-002-C2-2, RZ2004-00009, RZ2008-00006-C02-02 and RZ2008-00005-C02-02. We also thank the breeder associations who provided the biological samples. C. Ginja was supported by a grant from Fundação Portuguesa para a Ciência e a Tecnologia (Ref. SFRH/BD/13502/2003).

#### References

Barker JSF 1999. Conservation of livestock breed diversity. Animal Genetic Resources Information 25, 33–43.

Bennewitz J and Meuwissen THE 2004. Investigations on the selection of breeds for conservation of genetic variance. In 55th Annual Meeting of the European Association for Animal Production (ed. Y van der Honing), EAAP Book of Abstracts, Vol. 10. Bled, Slovenia.

Bennewitz J, Kantanen J, Tapio I, Li MH, Kalm E, Vilkki J, Ammosov I, Ivanova Z, Kiselyova T, Popov R and Meuwissen TH 2006. Estimation of breed contributions to present and future genetic diversity of 44 North Eurasian cattle breeds using core set diversity measures. Genetics Selection and Evolution 38, 201–220.

Boettcher PJ, Tixier-Boichard M, Toro MA, Simianer H, Eding H, Gandini G, Joost S, Garcia D, Colli L and Ajmone-Marsan P 2010. Objectives, criteria and methods for using molecular genetic data in priority setting for conservation of animal genetic resources. Animal Genetics 41 (suppl. 1), 64–77.

Caballero A and Toro MA 2002. Analysis of genetic diversity for the management of conserved subdivided populations. Conservation Genetics 3, 289–299.

Caballero A, Rodriguez-Ramilo ST, Avila V and Fernández J 2010. Management of genetic diversity of subdivided populations in conservation programmes. Conservation Genetics 11, 409–419.

Cañon J, Tupac-Yupanqui I, García-Atance MA, Cortés O, García D, Fernández J and Dunner S 2008. Genetic variation within the Lidia bovine breed. Animal Genetics 39, 439–445.

Cañon J, Alexandrino P, Bessa I, Carleos C, Carretero Y, Dunner S, Ferrand N, García D, Jordana J, Laloe D, Pereira A, Sanchez A and Moazami-Goudarzi K 2001. Genetic diversity measures of local European beef cattle breeds for conservation purposes. Genetics Selection and Evolution 33, 311–332.

Chaiwong N and Kinghorn BP 1999. Use of genetic markers to aid conservation decisions for groups of rare domestic breeds. Proceedings of the Association for the Advancement of Animal Breeding and Genetics 13, 365–368.

Cortes O, Tupac-Yupanqui I, Dunner S, Garcia-Atance MA, Garcia D, Fernandez J and Canon J 2008. Ancestral matrilineages and mitochondrial DNA diversity of the Lidia cattle breed. Animal Genetics 39, 649–654.

Cymbron T, Freeman AR, Isabel Malheiro M, Vigne JD and Bradley DG 2005. Microsatellite diversity suggests different histories for Mediterranean and Northern European cattle populations. Proceedings of the Royal Society of London, Series B, Biological sciences 272, 1837–1843.

Eding H and Meuwissen TH 2001. Marker-assisted estimates of between and within population kinships from genetic marker data for the construction of core sets in genetic conservation schemes. Journal of Animal Breeding and Genetics 118, 141–159. Eding H and Meuwissen THE 2003. Linear methods to estimate kinships from

genetic marker data for construction of core sets in genetic conservation schemes. Journal of Animal Breeding and Genetics 120, 289–302.

Eding H, Crooijmans RP, Groenen MA and Meuwissen TH 2002. Assessing the contribution of breeds to genetic diversity in conservation schemes. Genetics Selection and Evolution 34, 613–633.

European Cattle Genetic Diversity Consortium 2006. Marker-assisted conservation of European cattle breeds: an evaluation. Animal Genetics 37, 475–481.

Fabuel E, Barragan C, Silio L, Rodriguez MC and Toro MA 2004. Analysis of genetic diversity and conservation priorities in Iberian pigs based on microsatellite markers. Heredity 93, 104–113.

Falconer DS and Mackay TFC 1996. Introduction to quantitative genetics. Longman, Essex, UK.

Fernández J, Toro MA and Caballero A 2008. Management of subdivided populations in conservation programs: development of a novel dynamic system. Genetics 179, 683–692.

Food and Agriculture Organization (FAO) 1993. Secondary guidelines for development of National Farm Animal Genetic Resources Management Plans for measurement of domestic animal diversity (MoDAD). FAO, Original Working Group Report, Rome, Italy.

Food and Agriculture Organization (FAO) 2004. Measurement of domestic animal diversity – a review of recent diversity studies. FAO of the United Nations, Commission on Genetic Resources, Working Group on Animal Genetic Resources, Rome, Italy.

Garcia D, Corral N and Canon J 2005. Combining inter- and intrapopulation information with the Weitzman approach to diversity conservation. Journal of Heredity 96, 704–712.

Gibbs RA, Taylor JF, Van Tassell CP, Barendse W, Eversole KA, Gill CA, Green RD, Hamernik DL, Kappes SM, Lien S, Matukumalli LK, McEwan JC, Nazareth LV, Schnabel RD, Weinstock GM, Wheeler DA, Ajmone-Marsan P, Boettcher PJ, Caetano AR, Garcia JF, Hanotte O, Mariani P, Skow LC, Sonstegard TS, Williams JL, Diallo B, Hailemariam L, Martinez ML, Morris CA, Silva LO, Spelman RJ, Mulatu W, Zhao K, Abbey CA, Agaba M, Araujo FR, Bunch RJ, Burton J, Gorni C, Olivier H, Harrison BE, Luff B, Machado MA, Mwakaya J, Plastow G, Sim W, Smith T, Thomas MB, Valentini A, Williams P, Womack J, Woolliams JA, Liu Y, Qin X, Worley KC, Gao C, Jiang H, Moore SS, Ren Y, Song XZ, Bustamante CD, Hernandez RD, Muzny DM, Patil S, San Lucas A, Fu Q, Kent MP, Vega R, Matukumalli A, McWilliam S, Sclep G, Bryc K, Choi J, Gao H, Grefenstette JJ, Murdoch B, Stella A, Villa-Angulo R, Wright M, Aerts J, Jann O, Negrini R, Goddard ME, Hayes BJ, Bradley DG, Barbosa da Silva M, Lau LP, Liu GE, Lynn DJ, Panzitta F and Dodds KG 2009. Genome-wide survey of SNP variation uncovers the genetic structure of cattle breeds. Science 324, 528–532.

Ginja C, Telo Da Gama L and Penedo MC 2010a. Analysis of STR markers reveals high genetic structure in Portuguese native cattle. Journal of Heredity 101, 201–210.

Ginja C, Penedo MC, Sobral MF, Matos J, Borges C, Neves D, Rangel-Figueiredo T and Cravador A 2010b. Molecular genetic analysis of a cattle population to reconstitute the extinct Algarvia breed. Genetics Selection and Evolution 42, 18.

Ginja C, Penedo MC, Melucci L, Quiroz J, Martinez Lopez OR, Revidatti MA, Martinez-Martinez A, Delgado JV and Gama LT 2010c. Origins and genetic diversity of New World Creole cattle: inferences from mitochondrial and Y chromosome polymorphisms. Animal Genetics 41, 128–141.

Hayes B, He J, Moen T and Bennewitz J 2006. Use of molecular markers to maximise diversity of founder populations for aquaculture breeding programs. Aquaculture 255, 573–578.

Hodges J 1992. The management of global animal genetic resources. Food and Agriculture Organisation, Animal Production and Health Paper 104, Rome, Italy.

Martín-Burriel I, García-Muro E and Zaragoza P 1999. Genetic diversity analysis of six Spanish native cattle breeds using microsatellites. Animal Genetics 30, 177–182.

Martín-Burriel I, Rodellar C, Lenstra JA, Sanz A, Cons C, Osta R, Reta M, De Arguello S and Zaragoza P 2007. Genetic diversity and relationships of endangered Spanish cattle breeds. Journal of Heredity 98, 687–691.

Mateus JC, Eding H, Penedo MC and Rangel-Figueiredo MT 2004. Contributions of Portuguese cattle breeds to genetic diversity using marker-estimated kinships. Animal Genetics 35, 305–313.

Meuwissen T 2009. Towards consensus on how to measure neutral genetic diversity? Journal of Animal Breeding and Genetics 126, 333–334.

Ollivier L and Foulley JL 2005. Aggregate diversity: new approach combining within- and between-breed genetic diversity. Livestock Production Science 95, 247–254.

Perez-Figueroa A, Saura M, Fernández J, Toro MA and Caballero A 2009. METAPOP – a software for the management and analysis of subdivided populations in conservation programs. Conservation Genetics 10, 1097–1099.

Piyasatian N and Kinghorn BP 2003. Balancing genetic diversity, genetic merit and population viability in conservation programmes. Journal of Animal Breeding and Genetics 120, 137–149.

Reynolds J, Weir BS and Cockerham CC 1983. Estimation of the coancestry coefficient: basis for a short-term genetic distance. Genetics 105, 767–779.

Sánchez-Belda A 1984. Razas bovinas Españolas. Publicaciones de Extesión Agraria. MAPA, Madrid.

Scherf BD 2000. World watch list for domestic animal diversity. Food and Agriculture Organisation of the United Nations, Rome, Italy.

Tapio I, Varv S, Bennewitz J, Maleviciute J, Fimland E, Grislis Z, Meuwissen TH, Miceikiene I, Olsaker I, Viinalass H, Vilkki J and Kantanen J 2006. Prioritization for conservation of northern European cattle breeds based on analysis of microsatellite data. Conservation Biology 20, 1768–1779.

Toro MA and Caballero A 2005. Characterization and conservation of genetic diversity in subdivided populations. Philosophical Transactions of the Royal Society of London, Series B, Biological Sciences 360, 1367–1378.

Wang J and Hill WG 2000. Marker-assisted selection to increase effective population size by reducing Mendelian segregation variance. Genetics 154, 475–489.

Weitzman ML 1992. On diversity. The Quarterly Journal of Economics 107, 363–405.

Windig JJ and Engelsma KA 2010. Perspectives of genomics for genetic conservation of livestock. Conservation Genetics 11, 635–641.

Wright S 1951. The genetical structure of populations. Annals of Eugenics 15, 323–354.